

Effects of salicylic acid on physiological traits of myrtle seedlings in salt stress condition

¹Arsalan Shekarchian, ¹Vahid Etemad, ²Mohammad R. Bihamta,
³Mohammad H. Assareh

¹ Faculty of Natural Resources, University of Tehran, Karaj, Iran; ² Agronomy Department, Faculty of Agricultural Engineering and Technology, Karaj, Iran;
³ Plant Cell and Tissue Culture Research Institute, Karaj, Iran. Corresponding author:
A. Shekarchian, Arsalanshekar@yahoo.com

Abstract. Salicylic acid is known as an important signaling molecule that regulates plant reactions to environmental stresses. It plays a critical role in the regulation of physiological functions as non-enzymatic antioxidant. In this study we investigated the effects of different concentrations of salicylic acid (0, 50, 100 and 150 ppm) that were sprayed on one age seedlings of myrtle (*Myrtus communis* L.) with different levels of NaCl (0, 25, 50, and 100 mM) on activities of superoxide dismutase (SOD), guaiacol peroxidase (GPX), ascorbate peroxidase (APX), catalase (CAT), glycinebetaine (GB), proline, hydrogen peroxide (H₂O₂) and ionic leakage (ELI). Interactions of salicylic acid at 50 ppm with 50 mM salinity showed significantly increase in the CAT activity. GPX and APX were significantly higher in 50 mM salinity level with 50 and 100 ppm salicylic acid concentrations. The highest activity of SOD was observed in 50 mM of salt and 50 ppm of salicylic acid, indicated that 50 ppm had highest resistance against salt concentration. Proline in 50 mM salinity with 50 ppm salicylic acid showed the most significant increase in the 100 mM of salinity had a sharp decline in their interactions. GB had maximum amount in 25 mM salinity with all of salicylic acid treatments. Ionic leakage had significant interactions but was not significant with increasing salinity and salicylic acid treatments. The hydrogen peroxide showed the most significant increase in 100 mM salinity with 100 and 150 ppm salicylic acid levels respectively, and effectiveness of their interaction in 100 mM of salt with all treatments of salicylic acid except control.

Key Words: *Myrtus communis*, salinity, plant growth, salicylic acid.

Introduction. Soil salinity and alkalinity are main problems in almost all of irrigated areas and might be occur in agricultural fields and grasslands. In other words, all lands are capable to be salted. Soil salinity is one of the most serious problem attracts some scientists to overcome this obstruction by improving salt tolerant lines (Gholizadeh & Navabpour 2011).

Salinity stress reduces the plant growth and because of interruption in equivalence of essential elements absorption, water and oxidative tension, decrease the yield (Parida & Das 2005; Mollasiotis et al 2006). Stress induced reactive oxygen species (ROS) accumulation is counteracted by enzymatic antioxidant systems that include a variety of scavengers, such as superoxide dismutase (SOD), guaiacol peroxidase (GPX), ascorbate peroxidase (APX), catalase (CAT), and non-enzymatic low molecular metabolites, such as proline and sugar (Sarvajeet & Narendra 2010). The researches about impact of different tensions on grassland and forest plants are less than other fields in agriculture (Winicow 1998). Less proficiency of products such as subordinate forestial products, may be due to abiotic stresses (Shanker & Venkateswarlu 2011). One of the physiologic methods that in recent years, had been used for diminishing the environmental stresses on different plants, is application of tension extenuating materials (Yuan & Lin 2008). One of these materials is salicylic acid that is an important messenger molecule and it helps plant to response to the environmental tensions, and has a consequential role in adjustment of physiological activities in plant, such as a non-enzymatic antioxidant (Arfan et al 2007). Exogenous salicylic acid alters the activities of antioxidant enzymes and increases plant tolerance to abiotic stress by decreasing generation of ROS. It has been found that

salicylic acid has different effects on stress adaptation and damage development of plant that depend on plant species, concentration, method and time of salicylic acid application (Metwally et al 2003). Salicylic acid has obtained particular attention because of inducing protective effects on plants under NaCl salinity (Simaei et al 2011). Several studies have shown that the effect of cytotoxicity induced by salt stress can be ameliorated by the exogenous application of salicylic acid (Simaei et al 2012).

Myrtle (*Myrtus communis* L.) is an evergreen shrub that reaches a height of 3 meters. Myrtle has a considerable importance, given the role in semi-arid ecosystems in terms of soil conservation and prevent soil erosion and also its importance to use in multipurpose culture. Myrtle is a compatible plant in sub-humid areas of Iranian provinces included: Lorestan, Fars, Kerman, Sistan and Isfahan (Azadbakht 1999; Zargari 1996). The objectives of this work were to study the effect of salinity stress on the activities of antioxidant enzymes in myrtle and to examine whether the harmful effects of salinity stress can be offset by the exogenous application of salicylic acid.

Material and Method. In this study, seeds were collected from maternal plants growing in Jiroft, Kerman province of Iran. Pots were in green house of agricultural research and natural resources center of Kerman. During the emergence of seedlings, the weak seedlings were removed to have a suitable room in each pot.

This study was done using a factorial experiment in a randomized complete block design with four replications. Factors were included: four levels of salinity (0, 25, 50 and 100 mM) and four levels of salicylic acid (0, 50, 100 and 150 ppm).

Pot experiment done by 20-centimeter vases that were filled by sand, peat and field soil (proportion was 1:1:1). At the age of one year, salicylic acid was sprayed to pots at levels of: 0, 50, 100 and 150 ppm then salinity treatments at amounts of: 0, 25, 50 and 100 mM of sodium chloride were applied. The plants were irrigated by 3:1 Hoagland solution two times per week for two months and salicylic acid was sprayed on the plants every two weeks. Sodium chloride was added to 3:1 Hoagland solution (Tattini et al 2006). The amount of Glycinebetaine (GB) in dried leaves (third leaf from plant top) was measured (Grieve & Grattan 1983) and at a wavelength of 365 nm was read. Measurement of proline was done via spectrophotometry at $25 \pm 1^\circ\text{C}$ (Bates et al 1973) and at a wavelength of 520 nm was read. To measure the electrical conductivity of damage to membrane, ion leakage was assessed (Campos et al 2003).

Measurement of enzyme activity. CAT activity was measured by the destruction of H_2O_2 by CAT (Aebi 1984) and at a wavelength of 240 nm was read. APX activity was evaluated by method of Ranieri et al (2003) and at a wavelength of 290 nm was read. Measurement of GPX activity was done by method of Cakmak & Marschner (1988) and at a wavelength of 470 nm was read. Measuring the activity of superoxide dismutase was done photometric by method of Dhindsa et al (1981) and at a wavelength of 560 nm was read. The content of hydrogen peroxide was measured based on Loreto & Velikova (2001) and at a wavelength of 390 nm was read.

Statistical calculations. Analysis of variance was done by SAS (ver. 9.1) and the comparison of means was carried out using Duncan's multiple range test.

Results and Discussion. Table of analysis of variance (Table 1) showed that in this experiment salinity and salicylic acid treatments and interaction between salinity and salicylic acid at level of 1% for CAT were significant, the highest level of CAT enzyme was observed at 50 mM of salinity and then 25, 0 and 100 mM respectively. CAT enzyme activity at level of 25 and 50 mM of salinity, was more than control treatment, while this enzyme, only in control plants (at level of 0 salinity) there was more activity compared to 100 mM. There was a diminish in CAT enzyme activity at level of 100 mM that this is injurious to plant, because this decrease compared to control plants was significant. Assessing the main effect of salicylic acid showed that salicylic acid at 50 ppm improved CAT enzyme activity but it had a decrease trend at 100, 0 and 150 ppm respectively. The least CAT enzyme activity was observed at 150 ppm that this decline was significant.

Evaluation of interactions showed that at 100 mM salinity and 150 ppm salicylic acid, CAT enzyme had the least activity but the highest activity was observed at 50 mM salinity and 50 ppm salicylic acid (Figure 1).

Table 1

Analysis of variance mean squares of myrtle traits

Source	Df	CAT	GPX	SOD	APX	H ₂ O ₂	ELI	GB	Prolin
Replication	3	1.98	0.06	8.5	0.07	151.18	14.51	2.24	0.72
S	3	*26.34	*0.98	*185.38	*1.65	*49195.85	*2094.25	*304.35	*18.34
SA	3	*52.4	*0.65	*90.34	*1.42	*10474.58	52.41	*160.1	*27.04
S*SA	9	4.95	*0.18	*27.1	*0.2	*4113.01	28.59	17.25	*2.81
Error	3	2.41	0.04	7.83	0.05	1066.92	28.62	14.95	0.87
CV	45	20.97	25.87	25.72	29.55	13.42	24.14	25.3	22.22

p < 0.01 - *distinctly significant, S - salinity, SA - salicylic acid; Df - degrees of freedom; CAT - catalase; GPX - guaiacol peroxidase; SOD - superoxide dismutase; APX - ascorbate peroxidase; H₂O₂ - hydrogen peroxide; ELI - ionic leakage; GB - glycinebetaine.

GPX and APX enzymes showed same trend to different treatments. Table of analysis of variance showed that salinity and salicylic acid treatments and interaction between these two treatments showed a significant effect at probability 1%. The highest GPX and APX activity was at 50 mM salinity then at 25, 0 and 100 mM respectively. This plant at 100 mM salinity, compared to control, had less activity; that this decrease for GPX was significant but for APX was not. GPX enzyme was diminished at 100 mM salinity. Salicylic acid at 100 ppm, significantly encouraged the GPX and APX activity compared to control but at 150 ppm had opposite result and non-significantly decreased their activity. Interaction between salinity and salicylic acid for GPX and APX activity showed that these two enzymes at 50 mM salinity and 50 ppm salicylic acid, also at 50 mM salinity and 100 ppm salicylic acid had highest activity (Figures 2 and 3). Among different salinity levels, the highest GPX and APX activity was at 50 mM, then 25, 0 and 100 mM respectively. Myrtle at 100 mM salinity had less activity compared to control and this decrease for GPX was significant but for APX was not.

The highest level of SOD activity was seen at 50 mM salinity and then at 25, 0 and 100 mM respectively. SOD activity in control plants was higher than 100 mM and this difference was significant. Salicylic acid at level of 100 ppm showed highest level of SOD activity that it was significant. After that, the highest level of SOD activity was observed at 0, 50 and 150 ppm (Figure 4).

The highest level of prolin was observed at 50 mM salinity and then at 25, 0 and 100 mM. At 100 mM salinity, the prolin content was significantly reduced compared to control. The highest level of prolin content was seen at 50 ppm salicylic acid that this increase was significant. The least amount of prolin content was seen at 150 ppm and this decline was significant (Figure 5).

The highest level of glysin-betaein was observed at 25 mM salinity and then, at 0, 50 and 100 mM. In 100 mM salinity the amount of glysin-betaein was less than control plants and this slump was significant. With respect to the main effect of salicylic acid, at 50 ppm, glysin-betaein has highest level and this peak was significant; but difference between 0 and 100 ppm was not significant. The least amount of glysin-betaein was at 150 ppm that it was significant compared to other treatments except for control (Figure 6).

The highest quantity of ELI was at 100 mM salinity that it was noticeable compared to other treatments. Even though the least quantity of ELI was at 0 mM and its difference was significant compared to other treatments. Assessing the impact of salicylic acid showed that the least ELI was at 150 ppm that its difference between 0 ppm or 100 ppm was not significant (Figure 7).

The maximum amount of H₂O₂ was at 100 mM salinity and it was significantly different to other treatments. Among other different salinity treatments there was no significant difference. The main effect of salicylic acid on H₂O₂ showed that 150 ppm of

salicylic acid cause to highest synthesis of H_2O_2 and its difference from other treatments was significant; then had a dwindle trend at 100, 50 and 0 ppm, respectively (Figure 8).

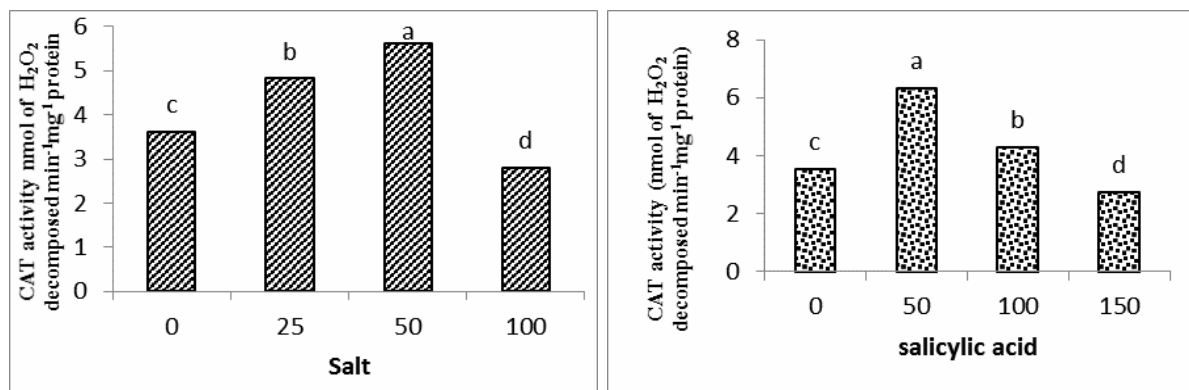


Figure 1. Effect of salinity and salicylic acid treatments on CAT activity.

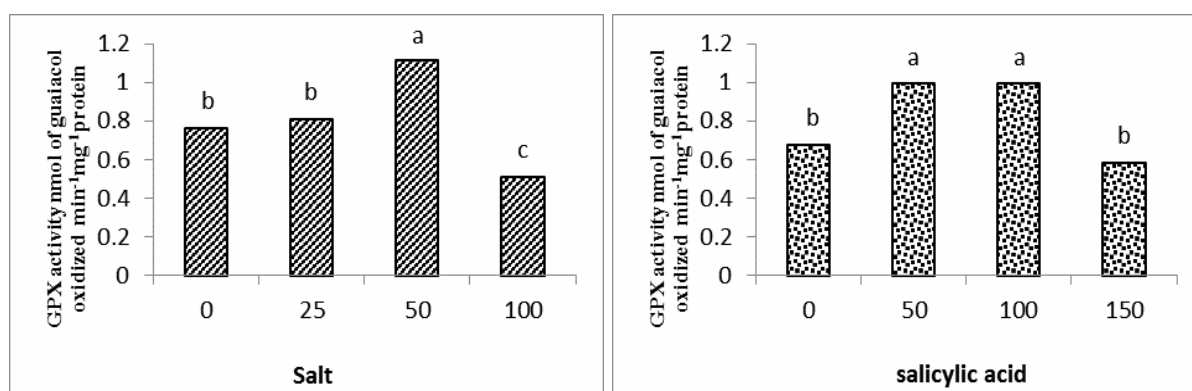


Figure 2. Effect of salinity and salicylic acid treatments on GPX activity.

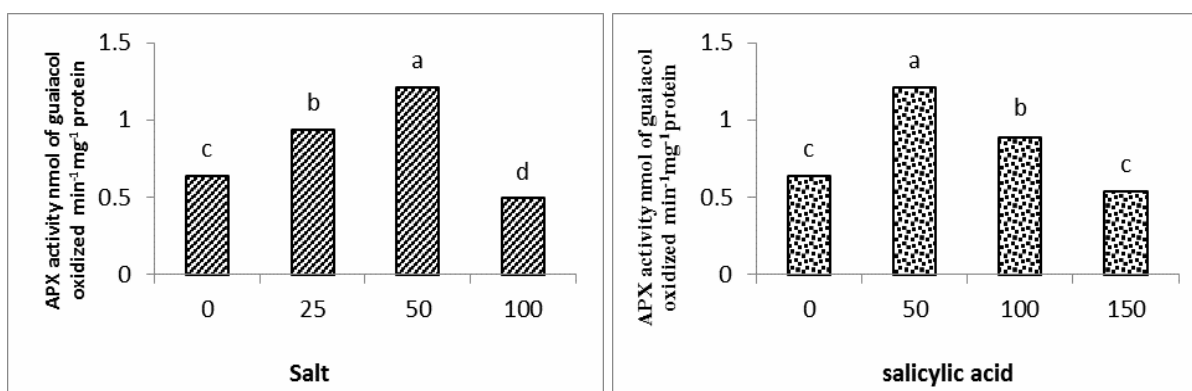


Figure 3. Effect of salinity and salicylic acid treatments on APX activity.

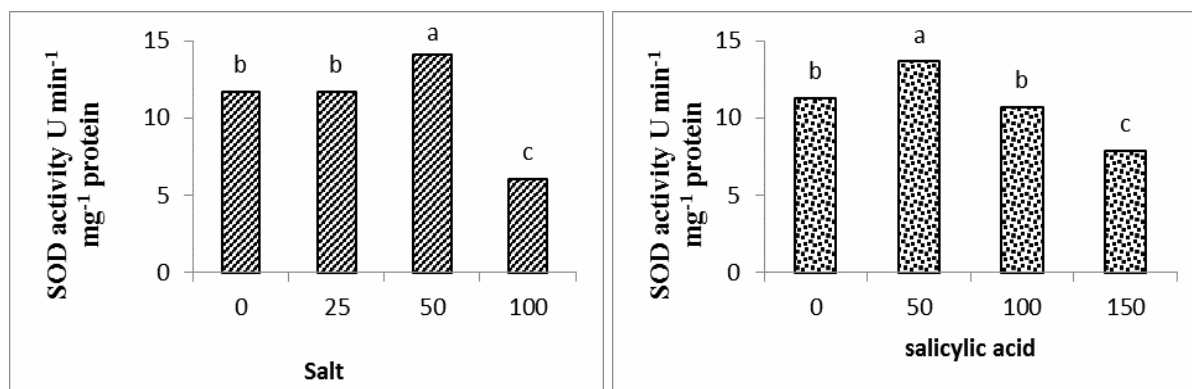


Figure 4. Effect of salinity and salicylic acid treatments on SOD activity.

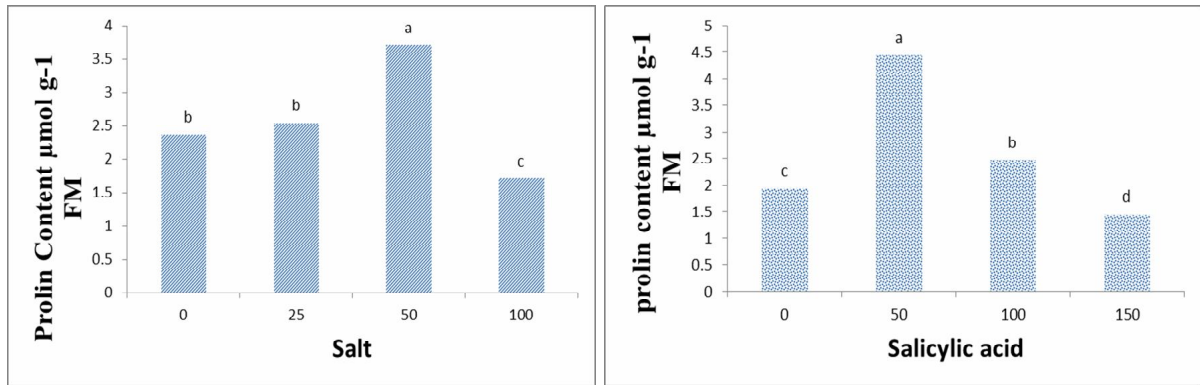


Figure 5. Effect of salinity and salicylic acid treatments on prolin content.

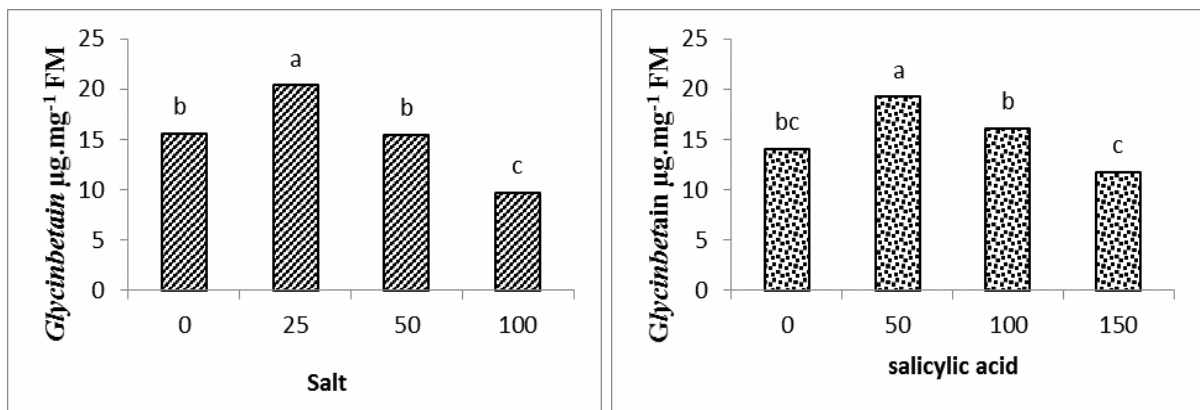


Figure 6. Effect of salinity and salicylic acid treatments on glycinebetaine activity.

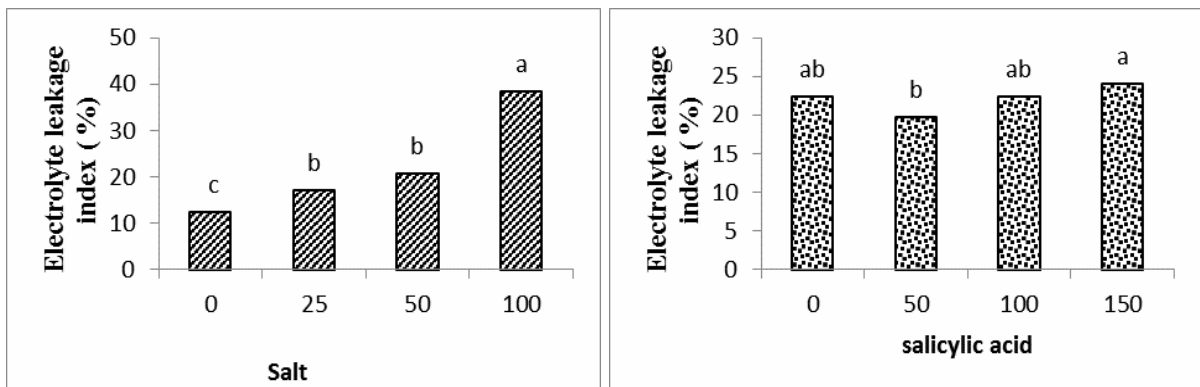


Figure 7. Effect of salinity and salicylic acid treatments on ionic leakage.

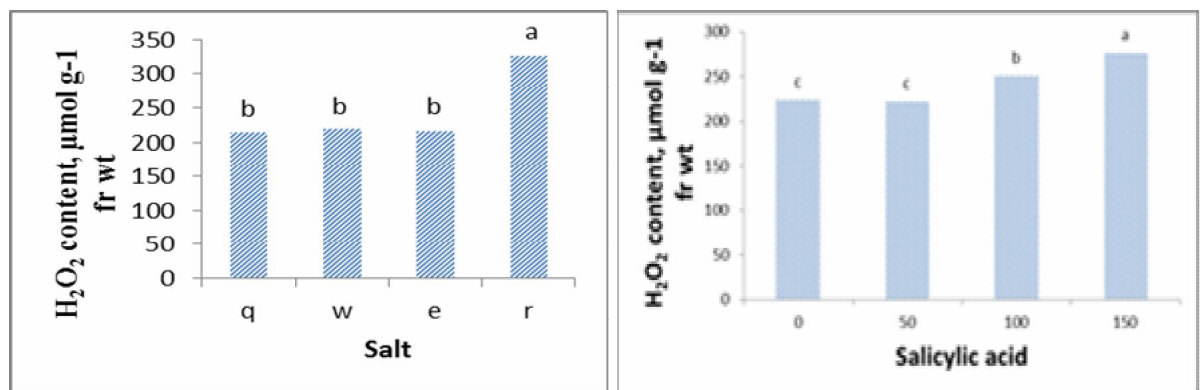


Figure 8. Effect of salinity and salicylic acid treatments on H₂O₂.

The decrease of CAT activity affected by 150 ppm salicylic acid was because of salinity tension conditions. Salicylic acid, inhibitor of CAT performance and by decrease of CAT activity and increase of H₂O₂ inside of plant (Horváth et al 2002). Also some reports acclaim that oxidative tensions deteriorate the CAT enzyme activity and escalate the amount of salicylic acid in rice, wheat, cucumber (Shim et al 2003) and in other plants (Sanchez-Casas & Klessig 1994). The improvement of effect between 50 ppm salicylic acid and 50 mM salinity, cause to increase antioxidant capacity of myrtle in direction of more production of CAT enzyme. Total consequence of salinity and salicylic acid impact and their interaction showed that 100 mM salinity and 150 ppm salicylic acid, reduce the CAT enzyme activity, hence plant tolerates this decline, because this reduction was not significant compared to control plants. But it shows a nosedive trend, that continues in higher concentrations of salinity and salicylic acid and at last, interrupts the physiologic status of plant. The highest activities of antioxidant system were obtained at 50 mM salinity and 50 ppm salicylic acid.

The least activity (significant) of GPX and APX was at 100 mM salinity and 150 ppm salicylic acid, that it can be representative of the injury to plant. The decrease of GPX enzyme at 100 mM salinity shows the ability of this level of salinity to reduce the antioxidant capacity. But GPX is not the main enzyme of H₂O₂ elimination and it has assistant contribution in H₂O₂ elimination, thus, this decrease of GPX cannot be seriously injurious to GPX. Leap of SOD activity cause to more production of H₂O₂ that it is poisonous to cell and should be immediately decomposed to water and oxygen by antioxidant defense system (Guo et al 2006). So, plant to get this purpose, utilizes other antioxidant enzymes to decompose the extra H₂O₂. Thus, it can be concluded that one of the reasons for increase of GPX activity in various levels of salinity tension is increase of SOD activity. Comparison of interaction between salinity and salicylic acid showed that addition of salicylic acid, improves the activity of peroxidase enzymes. It seems that higher concentrations of salicylic acid like 50 and 100 ppm besides of 50 mM salinity, cause to escalate the activity of GPX and APX enzymes that this result support research of Janda et al (1999). Eventually, results show that salicylic acid limitedly causes to remove of some disruptive impacts of salinity tensions especially prior to 50 mM.

Product of SOD activity (H₂O₂) is substrate of GPX and APX activity. So H₂O₂ can play as a signal to induce GPX and APX (Faize et al 2011). So, it is possible that a part of increase in GPX and APX activity, consequence of interaction between 50 mM salinity and 50 ppm salicylic acid also 50 mM salinity and 100 ppm salicylic acid, was due to rise of level of SOD enzyme. Decline of SOD at 100 mM salinity shows that this level of salinity could to reduce the antioxidant capacity of plant. The data of SOD confirms the data of CAT, GPX and APX, because interaction between 50 mM salinity and 50 ppm salicylic acid maximized the activity of each four antioxidant enzymes. Thus, it confirmed one of theories which acclaim that increase of CAT, GPX and APX enzymes is because of increase in SOD. H₂O₂ can play as signal for increasing of CAT, GPX and APX. At 50 mM salinity, by increasing the salicylic acid more than 50 ppm, the amount of SOD declined, especially there is the highest decrease of SOD at 100 mM salinity and 150 ppm salicylic acid and at this treatment the plant had injured, because the decline of antioxidant in treated plant was significant compared to control plant. These results showed that use of salicylic acid less than 50 ppm beside of 50 mM salinity, significantly increased the SOD activity compared to most of treatments. Salicylic acid improves the plant tolerance by increasing this enzyme and via this action, counteracts to tension. Impact of salicylic acid in maintenance of plant response in various oxidative tensions has been reported (Shirasu et al 1997).

Proline has different roles in osmotic tension included stability of proteins and cell membranes, sub-cell structures and protection of cell performances by remove the active oxygen elements (Chey & Li 2007). Results of this study showed a growth in prolin content in tensioned plants at 25 and 50 mM salinity; and supports the previous studies which acclaim this increase is due to defensive nature of prolin (El-Tayeb et al 2006). Research on black olives (Kouhifayegh et al 2013), rice (Weems et al 1999) and wheat (Wang et al 2007) under salinity tension, supports results of this study. Outcomes of this study are that apply the 50 mM salinity and 50 ppm salicylic acid cause to increase of

prolin accumulation. It can be assumed that treatment with salicylic acid lead to the prolin accumulation (Ashraf & Foolad 2007) which is consistent with the results of other researchers (Misra & Saxena 2009).

Glycinebetaine appears in plant by tension crisis and has high correlations with plant growth in dry and salty environments (Hanson et al 1985). Khan et al (2000) reported that by leaping the salinity, concentration of glycinebetaine in *Atriplex* spp. has increased. According to the present study, by increase of salinity tension levels to 25 mM, plant to maintenance the status of cells and tissues, could to increase the glycinebetaine; then it was steady at 50 mM salinity, and at 100 mM salinity showed a significant drop compared to control plants. Results showed that myrtle at initial stages of tension, used glycinebetaine as a mechanism to counteract the tension but at higher levels of tension, this plant utilized other mechanisms to counteract the tension. The impact of salicylic acid on accumulation of glycinebetaine has been proved (Hanson et al 1985; Hussain et al 2011; Khan et al 2014). In the present study, at 50 ppm salicylic acid, prolin was climbed significantly compared to other compounds. Effect of salicylic acid showed that the highest ELI observed at 150 ppm that it was significant and confirms results of Bastam et al (2013). Comparison of the mean of squares showed that salinity tension cause to increase the electrolyte leakage. Assessing the interaction effects showed the highest amount of ELI was observed at 100 mM salinity and different levels of salicylic acid, which indicates that decrease of antioxidant enzymes is due to injury to cell such as plasma membrane. Present study appeals that exactly the highest level of salinity and salicylic acid was injurious to plant, but the plant tolerated this injury and it was compensable. Because when amount of ELI gets to 50, from this amount is not more tolerable for plants (Lim et al 1998). While, at investigated treatments, the amount of ELI was less than 50.

Hydrogen peroxide is one of the effective factors in composition of lipid peroxidation (Sairam et al 2005; Prasad et al 1994). Also, hydrogen peroxide can contribute production of OH free radicals (another factor of oxidative tension) (Willekens et al 1995). Results confirm that higher level of salinity tension cause to higher amount of hydrogen peroxide (Figure 8). Increase H₂O₂ under tension may be due to the conversion of oxygen to hydrogen peroxide by SOD activity (Parida & Das 2005). In 100 mM salinity ELI was climbed, especially at interaction treatments between 100 mM salinity and all salicylic acid treatments (Figure 8). Increase of H₂O₂ cause to improve the ELI and decline of catalase, so increase of ELI, was considerably because of ROS invasion to cell plasma membrane of myrtle. Myrtle can tolerate 100 mM salinity but it was disruptive and the plant was injured because during the increase of salinity at the same time H₂O₂ has been increased, it means defense system of the plant could not maintain the level of H₂O₂ at low rate efficiently.

Conclusions. This study shows that myrtle recognized the salinity tension and responses to that. Salinity tension under 50 mM could activate the defense system but injury indexes (ELI and H₂O₂) show that 100 mM salinity damage to plant partly, that probably is tolerance threshold for myrtle. Also, salicylic acid at 50 ppm had a positive influence on physiologic system but at 150 ppm had negative effect on antioxidant system. The best treatment combination to improve defense system was 50 mM salinity and 50 ppm salicylic acid.

References

- Aebi H., 1984 Catalase in vitro. *Methods in Enzymology* 105:121–126.
- Arfan M., Athar H. R., Ashraf M., 2007 Does exogenous application of salicylic acid through the rooting medium modulate growth and photosynthetic capacity in two differently adapted spring wheat cultivars under saltstress? *Journal of Plant Physiology* 164:685-694.
- Ashraf M., Foolad M. R., 2007 Role of glycine betaine and proline in improving plant abiotic stress resistance. *Environmental and Experimental Botany* 59(2):206-216.

- Azadbakht M., 1999 [Classification of medical plants]. Teymourzade publications, Tabib Nashr [in Persian].
- Bastam N., Baninasab B., Ghobadi C., 2013 Improving salt tolerance by exogenous application of salicylic acid in seedlings of pistachio. *Plant Growth Regulation* 69(3):275-284.
- Bates L. S., Waldren R. P., Teare I. D., 1973 Rapid determination of free proline for water-stress studies. *Plant and Soil* 39:205–207.
- Campos P. S., Quartin V., Ramalho J. C., Nunes M. A., 2003 Electrolyte leakage and lipid degradation account for cold sensitivity in leaves of *Coffea* sp. plants. *Journal of Plant Physiology* 160:283–292.
- Cakmak I., Marschner H., 1988 Enhanced superoxide radical production in roots of zinc-deficient plants. *Journal of Experimental Botany* 39:1449-1460.
- Chey Y. L., Li Q. Z., 2007 Prediction of apoptosis protein subcellular location using improved hybrid approach and pseudo amino acid composition. *Journal of Theoretical Biology* 248(2):377-381.
- Dhindsa R. S., Plumb-Dhindsa P., Thorpe T. A., 1981 Leaf senescence: correlated with increased levels of membrane permeability and lipid peroxidation, and decreased levels of superoxide dismutase and catalase. *Journal of Experimental Botany* 32:93–101.
- El-Tayeb M., El-Enany A. E., Ahmed N. I., 2006 Salicylic acid-induced adaptive response to copper stress in sunflower (*Helianthus annuus* L.). *International Journal of Botany* 2:372-379.
- Faize M., Burgos L., Faize L., Piqueras A., Nicolas E., Barba-Espin G., Clemente-Moreno M. J., Alcobendas R., Artlip T., Hernandez J. A., 2011 Involvement of cytosolic ascorbate peroxidase and Cu/Zn-superoxide dismutase for improved tolerance against drought stress. *Journal of Experimental Botany* 62:2599-2613.
- Gholizadeh F., Navabpour S., 2011 Effect of salinity on morphological and physiological characteristics in correlation to selection of salt tolerance in rice (*Oryza sativa* L.). *International Journal of Agricultural Research* 6:780-788.
- Grieve C. M., Grattan S. R., 1983 Rapid assay for determination of water soluble quaternary ammonium compounds. *Plant Soil* 70:303-307.
- Guo Z., Ou W., Lu S., Zhong Q., 2006 Differential responses of antioxidative system to chilling and drought in four rice cultivars differing in sensitivity. *Plant Physiology and Biochemistry* 44(11-12):828-836.
- Hanson A. D., May A. M., Grumet R., Bode J., Jamieson G. C., Rhodes D., 1985 Betaine synthesis in chenopods: Location in chloroplasts: localization in chloroplasts. *Proceedings of the National Academy of Sciences of the USA* 82:3678-3682.
- Horváth E., Janda T., Szalai G., Páldi E., 2002 In vitro salicylic acid inhibition of catalase activity in maize: differences between the isozymes and a possible role in the induction of chilling tolerance. *Plant Science* 163:1129-1135.
- Hussain K., Navaz K., Majeed A., Ilyiaz U., Lin F., Ali K., Nisar M. F., 2011 Role of exogenous salicylic acid applications for salt tolerance in violet (*Viola odorata* L.). *Sarhad Journal of Agriculture* 27:171-175.
- Janda T., Szalai G., Tari I., Paldi E., 1999 Hydroponic treatment with salicylic acid decreases the effects of chilling injury in maize (*Zea mays* L.) plants. *Planta* 208:175-180.
- Khan M. A., Ungar I. A., Showalter A. M., 2000 Effects of salinity on growth, water relations and ion accumulation of the subtropical perennial halophyte, *Atriplex griffithii* var. *stocksii*. *Annals of Botany* 85(2):225-232.
- Khan M. I. R., Asgher M., Khan N. A., 2014 Alleviation of salt-induced photosynthesis and growth inhibition by salicylic acid involves glycinebetaine and ethylene in mungbean (*Vigna radiata* L.). *Plant Physiology and Biochemistry* 4:1-8.
- Kouhifayegh S. H., Hakimi M. H., Mosleh Arany A., Mirshamsi H. A., Kiani B., 2013 The effects of sodium nitroproside and salicylic acid on some physiological characteristics of *Melia azedarach* under salinity conditions. *Arid Biome Scientific and Research Journal* 3(2):62-71.

- Lim C. C., Arora R., Townsend E. C., 1998 Comparing Gompertz and Richard function to estimate freezing injury in *Rhododendron* using electrolyte leaking. *J Am Soc Hortic Sci* 123:346-252.
- Loreto F., Velikova V., 2001 Isoprene produced by leaves protects the photosynthetic apparatus against ozone damage, quenches ozone products, and reduces lipid peroxidation of cellular membranes. *Plant Physiology* 127:1781-1787.
- Metwally A., Finkemeier I., Georgi M., Dietz K. J., 2003 Salicylic acid alleviates the cadmium toxicity in barley seedlings. *Plant Physiology* 132(1):272-281.
- Misra N., Saxena P. 2009 Effect of salicylic acid on proline metabolism in lentil grown under salinity stress. *Plant Science* 177:181-189.
- Molassiotis A. N., Sotiropoulos T., Tanou G., Kofidis G., Diamantidis G., Therios I., 2006 Antioxidant and anatomical responses in shoot culture of the apple root stock MM 106 treated with NaCl, KCl, mannitol or sorbitol. *Biologia Plantarum* 50(1):61-68.
- Parida A. K., Das A. B., 2005 Salt tolerance and salinity effects on plants: a review. *Ecotoxicology and Environmental Safety* 60:324-349.
- Prasad T. K., Anderson M. D., Martin B. A., Stewart C. R., 1994 Evidence for chilling-induced oxidative stress in maize seedling and a regulatory role for hydrogen peroxide. *Plant Cell* 6:65-74.
- Ranieri A., Castagna A., Pacini J., Baldan B., Mensuali Sodi A., Soldatini G. F., 2003 Early production and scavenging of hydrogen peroxide in the apoplast of sun flower plants exposed to ozone. *Journal of Experimental Botany* 54(392):2529-2540.
- Sairam R. K., Srivastava G. C., Agarwal S., Meena R. C., 2005 Differences in antioxidant activity in response to salinity stress in tolerant and susceptible wheat genotypes. *Biologia Plantarum* 49:85-91.
- Sanchez-Casas P., Klessig D. F., 1994 A salicylic acid-binding activity and salicylic acid -inhibitable catalase activity are present in a variety of plant species. *Plant Physiology* 106(4):1675-1679.
- Sarvajeet S. G., Narendra T., 2010 Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plant. *Plant Physiology and Biochemistry* 48:909-930.
- Shanker A. K., Venkateswarlu B., 2011 Abiotic stress in plants - mechanisms and adaptations. InTech, ISBN 978-953-307-394-1, 440 pp.
- Shim I. S., Momose Y., Yamamoto A., Kim D. W., Usui K., 2003 Inhibition of catalase activity by oxidative stress and its relation to salicylic acid accumulation in plants. *Plant Growth Regulation* 39:285-292.
- Shirasu K., Nakajima H., Rajasekhar V. K., Dixon R. A., Lamb C., 1997 Salicylic acid potentiates an agonist-dependent gain control that amplifies pathogen signals in the activation of defense mechanisms. *Plant Cell* 9(2):261-270.
- Simaei M., Khavari-Nejad R. A., Bernard F., 2012 Exogenous application of salicylic acid and nitric oxide on the ionic contents and enzymatic activities in NaCl-stressed soybean plants. *American Journal of Plant Sciences* 3:1495-1503.
- Simaei M., Khavari-Nejad R. A., Saadatmand S., Bernard F., Fahimi H., 2011 Interactive effects of salicylic acid and nitric oxide on soybean plants under NaCl salinity. *Russian Journal of Plant Physiology* 58(5):783-390.
- Tattini M., Remorini D., Pinelli P., Agati G., Saracini E., Traversi M. L., Massai R., 2006 Morpho-anatomical, physiological and biochemical adjustments in response to root zone salinity stress and high solar radiation in two Mediterranean evergreen shrubs, *Myrtus communis* and *Pistacia lentiscus*. *New Phytologist* 170(4):779-794.
- Wang Z. Q., Yuan Y. Z., Ou J. Q., Lin Q. H., Zhang C. F., 2007 Glutamine synthetase and glutamate dehydrogenase contribute differentially to proline accumulation in leaves of wheat (*Triticum aestivum* L.) seedlings exposed to different salinity. *Journal of Plant Physiology* 164:695-701.
- Weems Y. S., Bridges P. J., LeaMaster B. R., Vincent D. L., Weems C. W., Sultana N., Ikeda T., Itoh R., 1999 Effect of NaCl salinity on photosynthesis and dry matter accumulation in developing rice grains. *Environmental and Experimental Botany* 42(3):211-220.

- Willekens H., Inzé D., Van Montagu M., Van Camp W., 1995 Catalases in plants. *Molecular Breeding* 1(3):207-228.
- Winicov I., 1998 New molecular approaches to improving salt tolerance in crop plants. *Annals of Botany* 82:703-710.
- Yuan S., Lin H. H., 2008 Role of salicylic acid in plant abiotic stress. *Zeitschrift fur Naturforschung C* 63: 313-320.
- Zargari A., 1996 Medicinal plants. Vol. 2. University of Tehran Publications.

Received: 01 May 2015. Accepted: 04 August 2015. Published online: 16 August 2015.

Authors:

Arsalan Shekarchian, Faculty of Natural Resources, University of Tehran, 315877821 Karaj, Iran, e-mail: Arsalanshekar@yahoo.com

Vahid Etemad, Faculty of Natural Resources, University of Tehran, 315877821 Karaj, Iran, e-mail: Vetemad@ut.ac.ir

Mohammad Reza Bihamta, Agronomy Department, Faculty of Agricultural Engineering and Technology, 315877821 Karaj, Iran, e-mail: Mrghanad@ut.ac.ir

Mohammad Hassan Assareh, Plant Cell and Tissue Culture Research Institute, 315351516 Karaj, Iran, e-mail: Asareh@gmail.com

This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

How to cite this article:

Shekarchian A., Etemad V., Bihamta M. R., Assareh M. H., 2016 Effects of salicylic acid on physiological traits of myrtle seedlings in salt stress condition. *AES Bioflux* 8(1): 14-23.